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## Recommended Citation

Onsare, R. S., Micoli, F., Lanzilao, L., Alfani, R., Okoro, C. K., Muigai, A. W., Revathi, G., Saul, A., Kariuki, S., MacLennan, C. A., Rondini, S. (2015). Relationship between Antibody Susceptibility and Lipopolysaccharide O-Antigen Characteristics of Invasive and Gastrointestinal Nontyphoidal Salmonellae Isolates from Kenya. *PLoS Neglected Tropical Diseases*, 9(3), 1-17.

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RESEARCH ARTICLE

# Relationship between Antibody Susceptibility and Lipopolysaccharide O-Antigen Characteristics of Invasive and Gastrointestinal Nontyphoidal *Salmonellae* Isolates from Kenya

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**OPEN ACCESS**

**Citation:** Onsare RS, Micoli F, Lanzilao L, Alfini R, Okoro C, Muigai AW, et al. (2015) Relationship between Antibody Susceptibility and Lipopolysaccharide O-Antigen Characteristics of Invasive and Gastrointestinal Nontyphoidal *Salmonellae* Isolates from Kenya. *PLoS Negl Trop Dis* 9(3): e0003573. doi:10.1371/journal.pntd.0003573

**Editor:** Stephen Baker, Oxford University Clinical Research Unit, VIETNAM

**Received:** August 21, 2014

**Accepted:** January 28, 2015

**Published:** March 4, 2015

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**Data Availability Statement:** All relevant data are within the paper.

**Funding:** This work was supported through a PhD scholarship grant from the European GENDRIVAX project (Grant Agreement no. PIAP-GA-2009-251522). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

## Abstract

### Background

Nontyphoidal *Salmonellae* (NTS) cause a large burden of invasive and gastrointestinal disease among young children in sub-Saharan Africa. No vaccine is currently available. Previous reports indicate the importance of the O-antigen of *Salmonella* lipopolysaccharide for virulence and resistance to antibody-mediated killing. We hypothesised that isolates with more O-antigen have increased resistance to antibody-mediated killing and are more likely to be invasive than gastrointestinal.

### Methodology/Principal Findings

We studied 192 NTS isolates (114 Typhimurium, 78 Enteritidis) from blood and stools, mostly from paediatric admissions in Kenya 2000–2011. Isolates were tested for susceptibility to antibody-mediated killing, using whole adult serum. O-antigen structural characteristics, including O-acetylation and glucosylation, were investigated. Overall, isolates were susceptible to antibody-mediated killing, but *S. Enteritidis* were less susceptible and expressed more O-antigen than Typhimurium ( $p < 0.0001$  for both comparisons). For *S. Typhimurium*, but not *Enteritidis*, O-antigen expression correlated with reduced sensitivity to killing ( $r = 0.29$ , 95% CI = 0.10–0.45,  $p = 0.002$ ). Both serovars expressed O-antigen populations ranging 21–33 kDa average molecular weight. O-antigen from most Typhimurium were O-acetylated on rhamnose and abequose residues, while *Enteritidis* O-antigen had low or no O-acetylation. Both Typhimurium and *Enteritidis* O-antigen were approximately

**Competing Interests:** FM, LL, RA, AS, CAM, SR are employees of Novartis. AS owns Novartis shares. This does not alter our adherence to all PLOS policies on sharing data and materials.

20%–50% glucosylated. Amount of *S. Typhimurium* O-antigen and O-antigen glucosylation level were inversely related. There was no clear association between clinical presentation and antibody susceptibility, O-antigen level or other O-antigen features.

## Conclusion/Significance

Kenyan *S. Typhimurium* and Enteritidis clinical isolates are susceptible to antibody-mediated killing, with degree of susceptibility varying with level of O-antigen for *S. Typhimurium*. This supports the development of an antibody-inducing vaccine against NTS for Africa. No clear differences were found in the phenotype of isolates from blood and stool, suggesting that the same isolates can cause invasive disease and gastroenteritis. Genome studies are required to understand whether invasive and gastrointestinal isolates differ at the genotypic level.

## Author Summary

Nontyphoidal *Salmonellae* (NTS) are an emerging major cause of invasive bacterial disease in African children aged less than 5 years and immunocompromised adults, with an estimated case fatality rate of 20–25%. NTS also cause diarrhoea, a killer of about 1.5 million young children annually, mainly in low- and middle-income countries. No vaccine against NTS is available, but improved understanding of the *Salmonella* bacteria that cause disease in Africa would help the development of new vaccines. The authors characterized a collection of 192 Kenyan NTS strains (114 *S. Typhimurium* and 78 *S. Enteritidis*) from blood and stool specimens. All strains could be killed to differing extents by antibodies present in the blood of healthy HIV-uninfected African adults, supporting the development of a vaccine that will induce protective antibodies when given to African children. Differences in killing by antibody were partly related to the amount of O-antigen on the bacterial surface. There were no clear distinction between stains causing invasive disease and diarrhoea, suggesting that the same strains may be capable of causing both forms of disease. Clarification of this will require genomic analysis.

## Introduction

NTS are a major but neglected cause of invasive disease (hence iNTS disease) in Africa [1–3]. *Salmonella enterica* serovars Typhimurium and Enteritidis account for nearly 80% of all human isolates reported globally [4]. While in developed countries, these predominantly cause a mild self-limiting gastroenteritis [5–7], in Africa they are responsible for bacteraemia, often associated with meningitis in young children, with incidence rates comparable to invasive *S. pneumoniae* disease [3]. The true burden of iNTS disease is uncertain due to the absence of a characteristic clinical presentation. Patients often present with nonspecific fever [8–10] and blood culture is necessary for diagnosis. Even where blood culture facilities are available, rapid clinical progression of NTS bacteraemia results in many patients dying before a microbiological diagnosis can be made [10]. No vaccine is available, and clinical management is made difficult by widespread multi-drug resistance and the need for late-generation expensive antibiotics [11–13].

In Kenya, iNTS disease is particularly frequent in rural areas [14], with incidence rates as high as 568/100,000 person-years [15]. A recent study from Western Kenya found an association between NTS diarrhoea and mortality in hospitalized children [16], indicating that NTS isolates in the region can cause fatal invasive and gastrointestinal disease, but it is currently unknown whether specific microbial phenotypic or genotypic characteristics are associated with each clinical presentations. Whole genome sequencing studies demonstrate that invasive African *S. Typhimurium* are genetically distinct from those in the rest of the world, and are characterized by a new sequence type, ST313, that has spread throughout the continent [17]. The ST313 pathovar is associated with genome degradation and pseudogene accumulation [13]. Equivalent studies are ongoing for *S. Enteritidis* which is more prevalent than *S. Typhimurium* in some parts of Africa [18,19], and globally [20]. Investigation of the relationship between genotypic and phenotypic features of *Typhimurium* isolates is ongoing and a systematic characterization of endemic *Typhimurium* and *Enteritidis* isolates from one endemic region is awaited.

Lipopolysaccharide (LPS) forms the outer layer of *Salmonella* and other Gram-negative bacteria, and is key to the interaction between *Salmonella* and its environment. The O-antigen chain (including core sugars, hereafter referred to as 'OAg') constitutes the outermost part of LPS [21]. In pathogenic bacteria such as *Salmonella*, LPS plays an important role in the interaction between the bacterium and its host and is a virulence factor required for colonization and resistance to antibody-mediated killing [22]. While the lipid A moiety is the predominant cause of the endotoxic effects of LPS, OAg is the most immunodominant portion of the molecule, and is responsible for serovar-specificity [23]. OAg protects bacteria from the environment and from serum complement [24–26], which can lead to bacterial killing by membrane attack complex formation. Rough strains of *Salmonella* that lack OAg are avirulent and succumb readily to complement-mediated killing [26]. The OAg structure plays a role in bacterial virulence, with longer OAg chains associated with increased complement and antibody resistance [27–29] and protection against other host antimicrobial factors [30].

In this study we analysed a bacterial collection of 114 *S. Typhimurium* and 78 *S. Enteritidis* isolates derived from patients, mostly children, presenting to three main Kenyan hospitals in Nairobi and from 31 healthy carriers related to these patients. Our primary objective was to characterise NTS isolates circulating in endemic areas, with the goal of identifying bacterial features associated with either invasive or gastrointestinal pathology. We hypothesised that invasive isolates would, by nature, be more resistant to survival in the blood than gastrointestinal isolates, and focused our analysis on the susceptibility of isolates to antibody-mediated killing and characterization of the OAg expressed by the various isolates. We investigated the total amount of OAg expressed by the various isolates and OAg specific characteristics such as molecular weight (MW), O-acetylation and glucosylation levels, parameters which can influence OAg immunogenicity. Additionally, we tested the sensitivity of the isolates to antibody killing, using whole human serum from HIV-uninfected adults [31]. The results obtained were compared with clinical presentation in order to identify possible associations.

## Materials and Methods

### Origin of bacterial isolates and serum samples

The study culture collection contained 192 archived NTS isolates (114 *S. Typhimurium* and 78 *S. Enteritidis*) collected between 2000 and 2011 at the Centre for Microbiology Research (CMR), Kenya Medical Research Institute (KEMRI) in Nairobi, Kenya. Study isolates were selected from the archived bacterial culture collection based on availability of unequivocal serovar identification and availability of clinical records and metadata. The isolates were stored in

Tryptic Soy Broth with 15% glycerol at -80°C. The isolates were from blood and stools with a few additional samples from urine (3 samples), cerebrospinal fluid (CSF, 2 samples) and environmental sources (2 samples). The isolates were mostly from children admitted to three main hospitals in Nairobi county, namely: Kenyatta National Referral hospital, Gertrude's Children's Hospital and Aga Khan University Hospital. This collection also included isolates from healthy carriers (31 samples). Before use in the present study, the archived isolates were re-cultured and the serotype was confirmed by antibody-based agglutination (Bio-Rad Laboratories, Inc., Hercules, CA, USA).

Based on the clinical sources and sample type, the NTS isolates were divided into four main groups. Those isolates from body fluids that are normally sterile such as blood and CSF were grouped as 'invasive', those from stool samples of patients were grouped as 'gastroenteric', those from stool samples of healthy individuals were grouped as 'healthy carriers' while those from soil and sewer water sources were grouped as 'environmental'. The three isolates from urine samples were considered clinically significant invasive sterile site isolates since *Salmonellae* are not part of the normal perineal skin flora and the isolates were from patients with symptoms of urinary tract infection.

Undiluted sera from ten healthy HIV-uninfected Malawian adults were used to generate a pooled serum to assess sensitivity to antibody-mediated killing of the *Salmonella* isolates. Each serum was tested prior to pooling to ensure that killing of the index *S. Typhimurium* ST313 strain, D23580, was within the 0.9 to 3.0 Log<sub>10</sub> range previously described for such sera [32]. Sera were handled at 4°C during the pooling process to ensure preservation of endogenous complement activity and frozen in aliquots at -80°C. Ethical approval for preparation of this serum was granted by the College of Medical Research and Ethics Committee, College of Medicine, University of Malawi [31].

### Serum Bactericidal Activity (SBA) assay

The study isolates were tested for their susceptibility/resistance to killing against the serum pool described above, using a protocol involving whole serum and endogenous complement activity as previously described [26]. Briefly, 5 µl washed bacteria at 2 h log-growth phase at an OD of approximately 0.2 (with shaking at 180 rpm) was added to 45 µl undiluted serum at a final bacterial concentration of 1×10<sup>6</sup> colony forming units, CFU/ml and incubated at 37°C with the number of viable bacteria count determined by serial dilution on Luria Bertani (LB) agar after 0 and 180 min. As a negative control, reference sera were heat-inactivated at 56°C for 45 min to inactivate endogenous complement and included in each SBA experiment. Additional internal controls included testing the representative endemic African *S. Typhimurium* isolate D23580, previously shown to be susceptible to serum killing [26] and hyperimmune mouse serum containing high levels of anti-OAg antibodies [33]. Susceptibility to serum killing was determined as any reduction in viable bacterial count compared with the initial *Salmonella* concentration.

### O-antigen extraction and quantification

OAg extraction was performed by acid hydrolysis [34]. Bacterial isolates were grown overnight in LB medium. As OAg expression can be influenced by growth conditions, identical conditions were used for the growth of all strains [29]. The bacterial OD was measured and the bacterial cultures were concentrated in PBS to OD: 35. Acetic acid (2% v/v) was then added to the concentrated growth bacterial culture (pH 3), which were incubated for 3h at 100°C. The reaction was stopped by the addition of 14% ammonium hydroxide, increasing the pH to around 6. Bacterial debris was removed by centrifugation and OAg in the supernatant was 0.22 µm

microfiltered and desalted by HiTrap Desalting 5 ml columns (GE Healthcare, UK). Phenol sulphuric acid assay, using glucose as standard, was used for quantification of OAg content [34,35].

### <sup>1</sup>H Nuclear Magnetic Resonance Spectroscopy (<sup>1</sup>H NMR)

<sup>1</sup>H NMR analysis was performed as a confirmation of the identity of the OAg samples and to verify the presence of O-acetyl groups along the OAg chain as previously described [34].

### High Performance Anion Exchange Chromatography with Pulsed Amperometric Detection (HPAEC-PAD)

Glucosylation level of OAg samples was estimated by HPAEC—PAD after acid hydrolysis of the OAg to release the monosaccharides constituting the sugar chain as described previously. Commercial glucose was used for building the calibration curve (0.5–10 µg/ml). Glucose (Glc) level is calculated as molar ratio of Glc to rhamnose (Rha), sugar present in each OAg repeating unit. The analysis was also used as confirmation of the OAg samples identity verifying the correct monosaccharides ratios in the repeating unit [34,36].

### Size-exclusion High-Performance Liquid Chromatography (HPLC-SEC)

HPLC—SEC analysis was used to estimate the molecular size distribution of OAg populations [34,36]. Samples were run, without pre-treatment, on a TSK gel G3000 PWXL column (30 cm x 7.8 mm; particle size 7 µm; cod. 808021) with a TSK gel PWXL guard column (4.0 cm x 6.0 mm; particle size 12 µm; cod. 808033) (Tosoh Bioscience, Tokyo, Japan). The mobile phase was 0.1 M NaCl, 0.1 M NaH<sub>2</sub>PO<sub>4</sub>, 5% CH<sub>3</sub>CN, pH 7.2, at the flow rate of 0.5 ml/min (isocratic method for 30 min). OAg peaks were detected by differential refractive index (dRI). Void and bed volume calibration was performed with λ-DNA (λ-DNA molecular weight (MW) Marker III 0.12–21.2 kb; Roche) and sodium azide (Merck, New Jersey, USA), respectively. OAg average MW was estimated on standard dextrans (Sigma) calibration curve.

### Statistical analyses

Serum susceptibility and OAg amounts of the two *Salmonella* serovars examined (*S. Typhimurium* and *S. Enteritidis*) was compared using the Mann-Whitney U test. Possible correlations (i.e. OAg amount and serum susceptibility, glucosylation levels and OAg production) were analysed by Spearman rank.

## Results

### NTS bacterial collection

Of the 192 NTS isolates in the study, 114 (59%) were *S. Typhimurium* and 78 (41%) were *S. Enteritidis*. This study collection included isolates from blood, stools (patients and healthy contacts), CSF, urine and environmental/animal sources as described in Table 1 (see S1a-b Table for supporting information). The isolates were obtained from hospitals based in Nairobi, an urban setting. Clinical information on patients' age was available for 67% of cases (73 with *S. Typhimurium*, 57 with *S. Enteritidis*). The age ranged from 1 month to 59 years for *S. Typhimurium* isolates and 0 months to 64 years for *S. Enteritidis* isolates, the majority (65%, 85 isolates) of these patients being below 5 years (53 *S. Typhimurium*, 32 *S. Enteritidis*). Overall, the median age of subjects from whom *S. Typhimurium* and *S. Enteritidis* strains were isolated was 2 and 3.5 years, respectively. Ages were not available for the healthy carriers.

**Table 1. Number of study NTS isolates by serovar and origin.**

	S. Typhimurium (% of total S. Typhimurium, n = 114)								S. Enteritidis (% of total S. Enteritidis, n = 78)				Total (% of total NTS)
	blood	Stool	blood & stool	urine	CSF	Soil	Sewer water	ND	blood	Stool	blood & stool	ND	
Invasive	48 (42%)	–	7 (6%)	3 (3%)	2 (2%)	–	–	–	22 (28%)	–	5 (6%)	–	87 (45%)
Gastrointestinal	–	24 (21%)	–	–	–	–	–	–	–	32 (41%)	–	–	56 (29%)
Healthy carriers	–	21 (18%)	–	–	–	–	–	–	–	10 (13%)	–	–	31 (16%)
Environmental	–	–	–	–	–	1 (1%)	1 (1%)	–	–	–	–	–	2 (1%)
ND	–	–	–	–	–	–	–	7 (6%)	–	–	–	9 (12%)	16 (8%)
<b>Total</b>	<b>114</b>								<b>78</b>				<b>192</b>

ND = Isolates for which no documentation on clinical form was available; n = Total number Gastrointestinal and healthy carrier isolates were derived from stool samples.

doi:10.1371/journal.pntd.0003573.t001

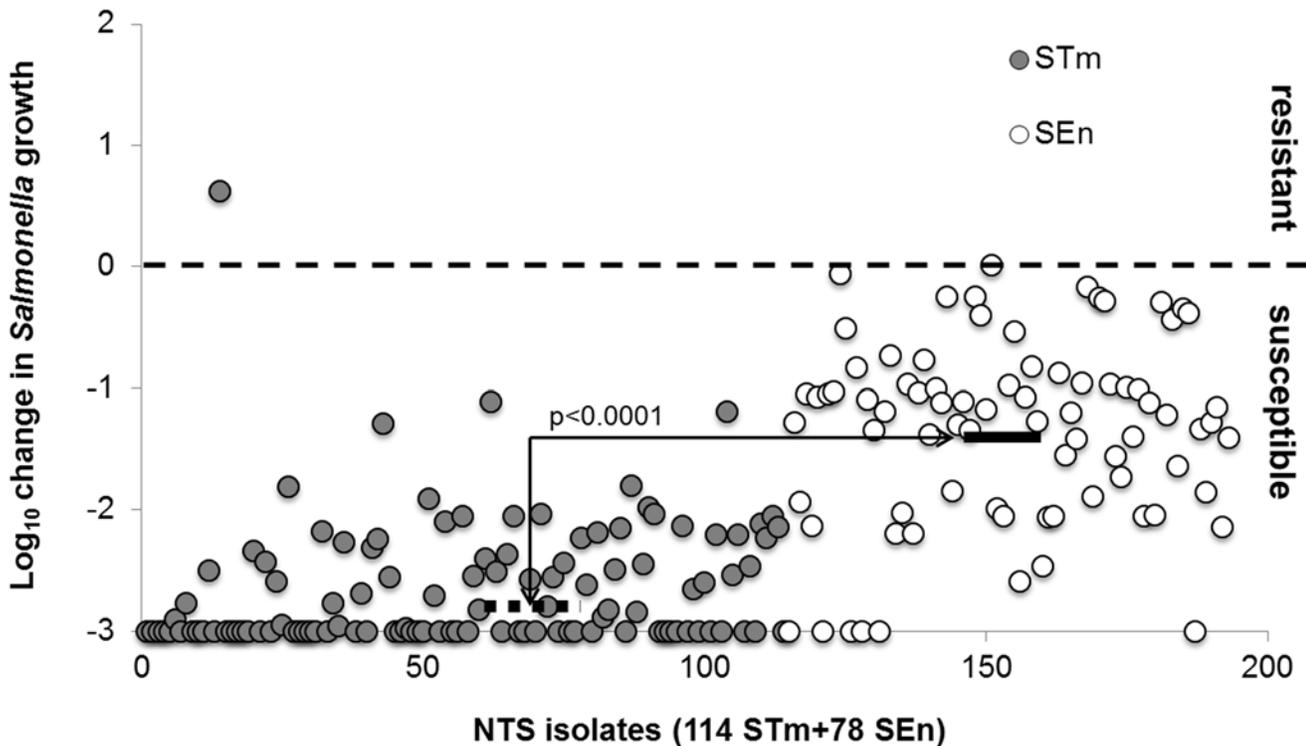
### SBA results

SBA results using the serum pool demonstrated a marked difference in susceptibility to antibody-mediated killing among individual isolates and a clear difference in susceptibility between *S. Typhimurium* and *S. Enteritidis* isolates (Fig. 1, and see S2a and S2b Table for supporting information). We have previously demonstrated that this killing is through the antibody-dependent complement-mediated mechanism and requires the presence of specific antibodies and intact complement function [26]. We have also demonstrated that the control sera used in this assay from healthy Malawian adults contains abundant antibody levels to NTS [32]. All except one *S. Typhimurium* isolate (0.9%, 1/114) underwent a reduction in viable bacterial counts over the 180 minute time course of the assay, and so were designated ‘sensitive’ to antibody-mediated killing. Overall median Log<sub>10</sub> reduction in viable bacteria after 180 minutes was 2.9 for *S. Typhimurium*. Indeed 50% (57/114) *S. Typhimurium* were highly susceptible to killing, undergoing a 3.0 Log<sub>10</sub> reduction. In contrast *S. Enteritidis* as a group were less susceptible to antibody-mediated killing than *S. Typhimurium*, being killed by a median of 1.2 Log<sub>10</sub> (p<0.0001, Mann Whitney test). However, all *S. Enteritidis* were sensitive to killing. A small proportion (10.3%, 6/78) was killed by a full 3.0 Log<sub>10</sub>.

### OAg characterization

OAg was extracted from all 192 NTS study isolates normalized to the same final OD and total amount was quantified by phenol sulphuric acid assay [34]. NTS isolates produced a range of OAg amounts, with low (<10 µg/ml/OD) and high producers (>25 µg/ml/OD). On average, *S. Enteritidis* isolates expressed more OAg than *S. Typhimurium*, with 70.9% of the isolates producing more than 15 µg/ml/OD OAg compared to only 42.1% *S. Typhimurium*. Median OAg production for *S. Enteritidis* isolates was 16.8 µg/ml/OD compared to 14.4 µg/ml/OD of *S. Typhimurium* (p<0.0001 by Mann Whitney test) (Fig. 2). All study isolates contained one single main OAg population with average MW of 21–33 kDa (Fig. 3a); <5% isolates contained OAg population with average MW < 6 kDa.

NMR analysis on a set of 118 samples (73 *S. Typhimurium* and 45 *S. Enteritidis*) showed that only 6.8% of *S. Typhimurium* OAg were not O-acetylated, compared with 35.6% and



**Fig 1. SBA results of STm and SEN isolates with Malawi serum pool.** Bacterial colony forming units (CFU) were counted at time 0 (T0) and after 3 h (T180) incubation of bacteria in 45  $\mu$ L undiluted serum (final bacterial concentration  $1 \times 10^6$  CFU/ml). Killing/growth was determined by  $\text{Log}_{10}$  (CFU at T180) -  $\text{Log}_{10}$  (CFU at T0). Strains that could grow ( $\text{Log}_{10}$  change  $> 0$ ) were considered “resistant”, strains that were killed ( $\text{Log}_{10}$  change  $< 0$ ) were considered susceptible. Dashed/continuous line indicates median  $\text{Log}_{10}$  change of STm (-2.9) and SEN (-1.2) isolates, respectively.

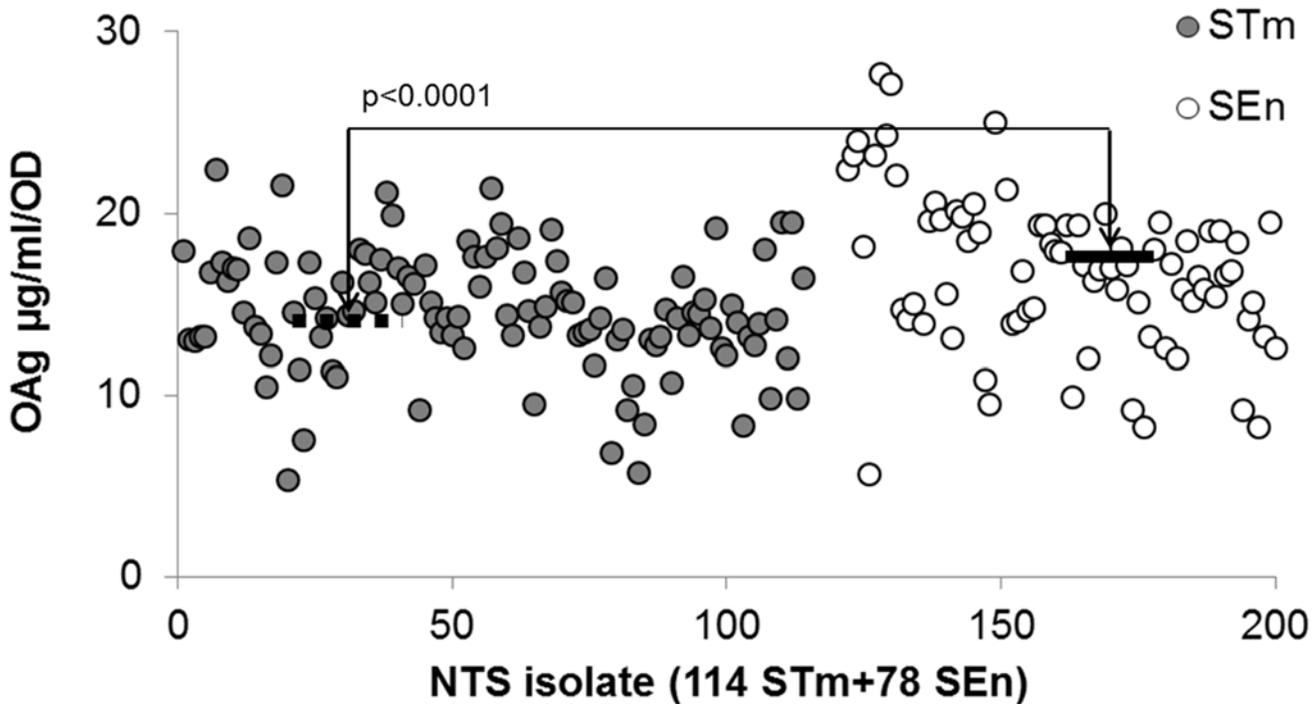
doi:10.1371/journal.pntd.0003573.g001

42.2% of *S. Enteritidis* OAg with no or traces of O-acetyl groups, respectively. For *S. Typhimurium*, OAg O-acetylation was detected on both rhamnose (Rha) and abequeose (Abe) sugar residues in the majority of isolates (63%), but also on Rha only (9.6%) and Abe only (17.8%) (Table 2; Fig. 3b and c; see S3a Table for supporting information). HPAEC-PAD was done on a subset of 91 isolates (55 *S. Typhimurium* and 36 SEN) and showed that for both serovars, the majority of strains (72.2%) expressed OAg with 20–50% glucosylation levels (Fig. 3d, see S3b Table for supporting information).

### Analysis of associations between clinical presentation and bacterial characteristics

SBA results, OAg features and clinical presentation were compared. As mentioned above, there was a clear inter-serovar difference, with *S. Enteritidis* isolates having more OAg than *S. Typhimurium* and being more resistant to antibody-mediated killing than *S. Typhimurium*. Among *S. Typhimurium* isolates, there was a correlation between amount of OAg and resistance to antibody-mediated killing (Spearman  $r = 0.29$ , 95% CI 0.10 to 0.45,  $p = 0.002$ ) in accordance with our initial hypothesis, but this was not present for *S. Enteritidis* (Fig. 4).

No correlation was found between NTS clinical presentation (invasive, gastrointestinal and carrier) and either antibody susceptibility or polysaccharide production, for either *S. Typhimurium* or *S. Enteritidis* isolates (Fig. 5). There was no clear correlation between O-acetylation levels/position and glucosylation levels with clinical presentation, antibody susceptibility and OAg production. For *S. Typhimurium*, we found an association between



**Fig 2. OAg production of *S. Typhimurium* (STm) and *S. Enteritidis* (SEn) isolates after growth and normalization to the same final OD: 35.** Dashed/continuous line indicates median OAg production of *S. Typhimurium* (STm) (14.4 µg/ml/OD) and *S. Enteritidis* (SEn) (16.8 µg/ml/OD) isolates, respectively.

doi:10.1371/journal.pntd.0003573.g002

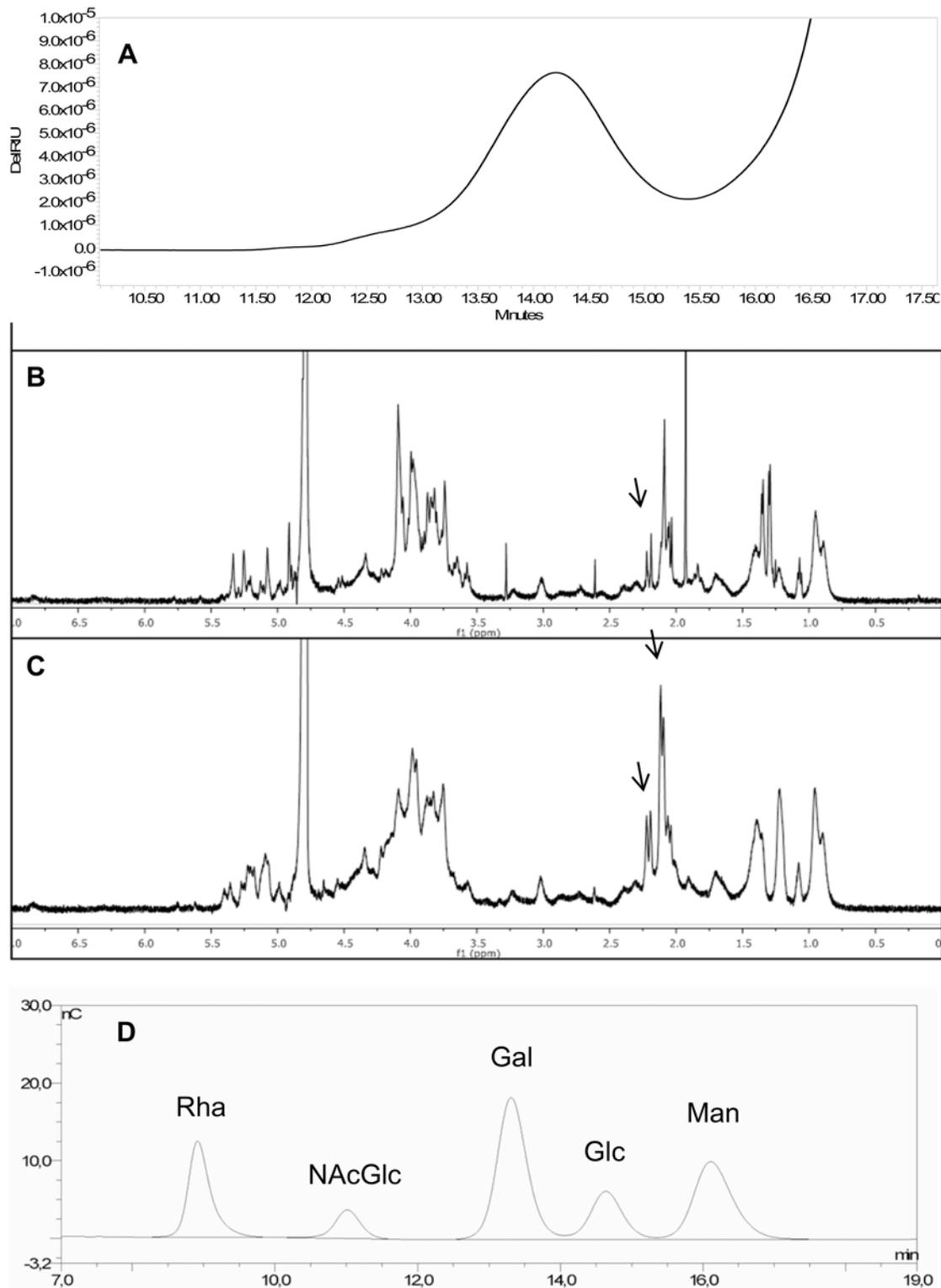
lower glucosylation levels and higher OAg production (Spearman  $r = 0.51$ , 95% CI-0.69 to -0.27,  $p$  value < 0.001) (Fig. 6).

## Discussion

NTS infections are a major problem in sub-Saharan Africa and can either present as bacteraemia, with symptoms of gastroenteritis in under half of patients, or as diarrheal disease without bloodstream infection [1,11,37]. Both presentations can be life-threatening, with case fatality rates of 20–25% for iNTS bacteraemia in children [1,2,8,10] and up to 50% in HIV-infected adults [9], and a recognized association between NTS diarrhoea with mortality [16]. No vaccine is currently available.

The main finding of the study is that all isolates studied, apart from one *S. Typhimurium*, were susceptible to antibody-dependent complement-mediated killing, using a serum pool from an endemic region. Previously it has been shown that the antibody response specific to NTS is acquired with age, corresponding with a decline in cases of NTS bacteraemia [26,38]. The serum pool we used contained high levels of anti-OAg (both *S. Typhimurium* and *S. Enteritidis*) antibodies as expected among healthy adults in a *Salmonella* endemic region of sub-Saharan Africa such as Kenya, Malawi or Tanzania [32]. The study therefore confirms the bactericidal activity of anti-*Salmonella* antibodies against the vast majority of endemic NTS isolates in the current study. These findings support an important role for antibodies in protective vaccines against iNTS disease [31,33,39].

Another key finding is the difference in relationship between susceptibility to antibody-mediated killing and levels of OAg expression for the two *Salmonella* serovars studied. According to our study hypothesis, decreased susceptibility of *S. Enteritidis* compared with *S. Typhimurium* could be the result of higher levels of OAg, not excluding the contributions of other



**Fig 3.** a) HPLC-SEC profile of *S. Typhimurium* Ke212 OAg (similar profiles obtained for all NTS strains); b)  $^1\text{H}$  NMR of *S. Enteritidis* Ke189 OAg and c) *S. Typhimurium* Ke004 OAg (arrows indicate O-acetyl groups); d) HPAEC-PAD profile of *S. Typhimurium* Ke012 OAg sample indicating typical sugars of the repeating unit (Rha, Gal, Glc and Man) and of the core region (Gal, Glc and NAcGlc) (similar profiles obtained for all NTS strains).

doi:10.1371/journal.pntd.0003573.g003

**Table 2. Proportion distribution in percentages of O-acetylation and glucosylation levels among NTS isolates.**

	O-Acetylation (n = 73 <i>S. Typhimurium</i> , 44 <i>S. Enteritidis</i> )					Glucosylation (n = 55 <i>S. Typhimurium</i> , 36 <i>S. Enteritidis</i> )		
	% strains with O-acetyl groups			% strains with traces or no O-acetyl groups		% strains with glucosylation level		
	On both Rha & Abe	On Abe only	On Rha only	With traces of O-acetyl groups	No O-acetyl groups	<20%	20–50%	>50%
<b><i>S. Typhimurium</i></b>	63.0	17.8	9.6	3.0	6.8	29.1	56.4	14.5
<b><i>S. Enteritidis</i></b>	22.2			42.2	35.6	13.9	72.2	13.9

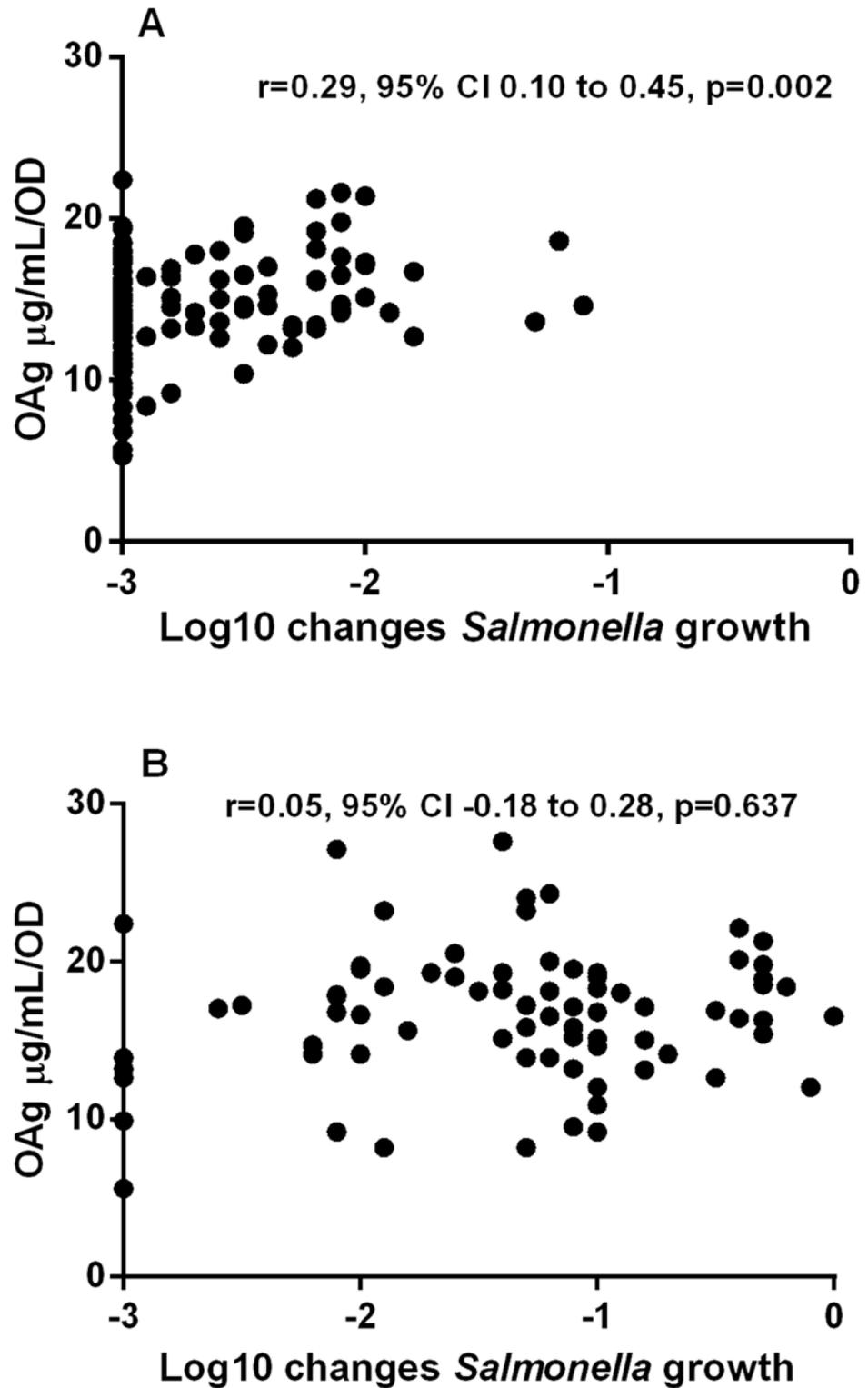
n = Total number; Rha = rhamnose; Abe = abequeose

doi:10.1371/journal.pntd.0003573.t002

pathogen features, not investigated here. A previous report from Malaysia found that *S. Enteritidis* isolates were more often isolated from the blood than stool of patients compared with other *Salmonella* serovars [40], but the reasons for this were not explored. Considering that OAg length and density play a role in pathogen virulence [27], increased levels of OAg expression may enhance the ability of *S. Enteritidis* isolates to evade host first line defence mechanisms and gain access to the bloodstream. It has also been reported that, when exposed to serum, *S. Typhimurium* upregulates genes for increased production of very long chain OAg to escape complement-dependent killing [28]. Our finding of a correlation between OAg expression and resistance to antibody-mediated killing among *S. Typhimurium* isolates is consistent with the hypothesis that high OAg expression protects against such killing. It is also well known that strains lacking OAg are rapidly eliminated from the host [41,42], and there were no such isolates in our collection. However, the lack of correlation between OAg expressed by *S. Enteritidis* isolates and susceptibility to antibody-mediated killing does not support our hypothesis for this serovar concerning OAg levels and resistance to antibody killing. This finding suggests that other inherent factors of *Salmonella*, particularly *S. Enteritidis*, have a role in determining loss of sensitivity. In other words, quantity of OAg is more important for resistance to antibody-killing of *S. Typhimurium* than *S. Enteritidis*. Further studies are required to better understand the factors contributing to the inherent resistance of *S. Enteritidis*.

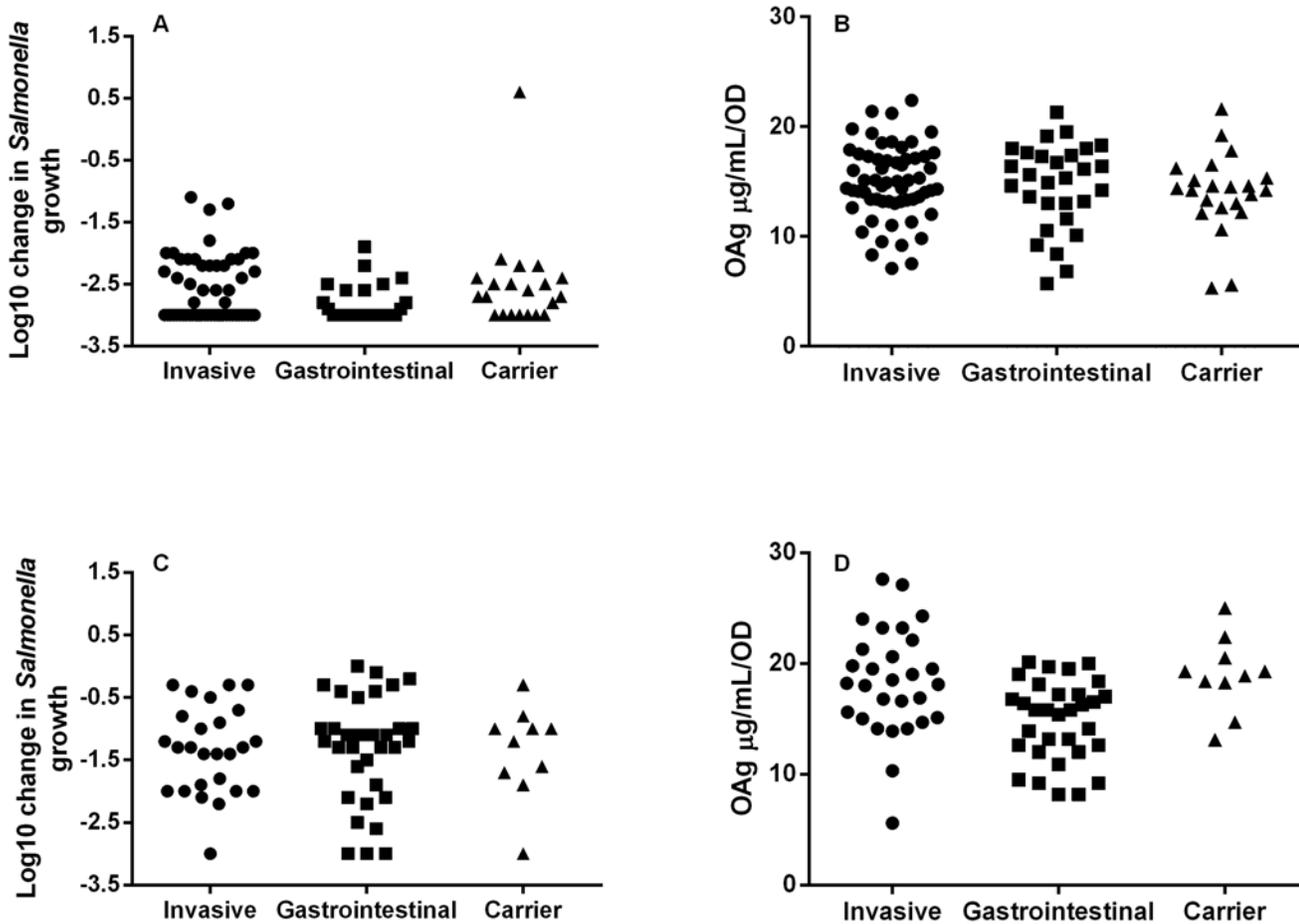
In relation to OAg fine specificity (O-acetylation) of the isolates in our collection, *S. Typhimurium* were more heterogeneous than *S. Enteritidis*. OAg O-acetylation occurred in almost all *S. Typhimurium* isolates, and most often at two different sites: C-2 Abe (known to provide factor O:5 specificity) and C-2/C-3 Rha. Rha O-acetylation was first reported by Wollin et al [43] and we have previously reported this as part of the characterisation of an invasive Malawian *S. Typhimurium* isolate [44]. The extent to which this additional O-acetylation was widespread in our collection suggests a possible role in pathogenesis that should be investigated further. Moreover, the fact that we could also identify *S. Typhimurium* isolates with either Abe- or Rha-O-acetylation only, shows that O-acetylation at both sites can occur independently. A *S. Typhimurium* OAg-based vaccine for Africa would need to induce antibodies capable of binding to both OAg specificities. In contrast, O-acetylation of *S. Enteritidis* OAg was not as common, with more than two-thirds of the isolates lacking or having very low levels of O-acetylation.

Glucosylation is another OAg modification for which variation has been described for *S. Typhimurium* and can occur on the O:12 antigen galactose (C4 position) generating the 12–2 variant, and on the O:1 antigen galactose (C6 position) (phage-determined) [45]. It has been



**Fig 4. Correlation of SBA results and OAg production levels in (a) *S. Typhimurium* isolates ( $p = 0.002$ ) and (b) *S. Enteritidis* isolates ( $p = 0.637$ ).**  $r$  is the Spearman correlation coefficient. CI is confidence interval.

doi:10.1371/journal.pntd.0003573.g004

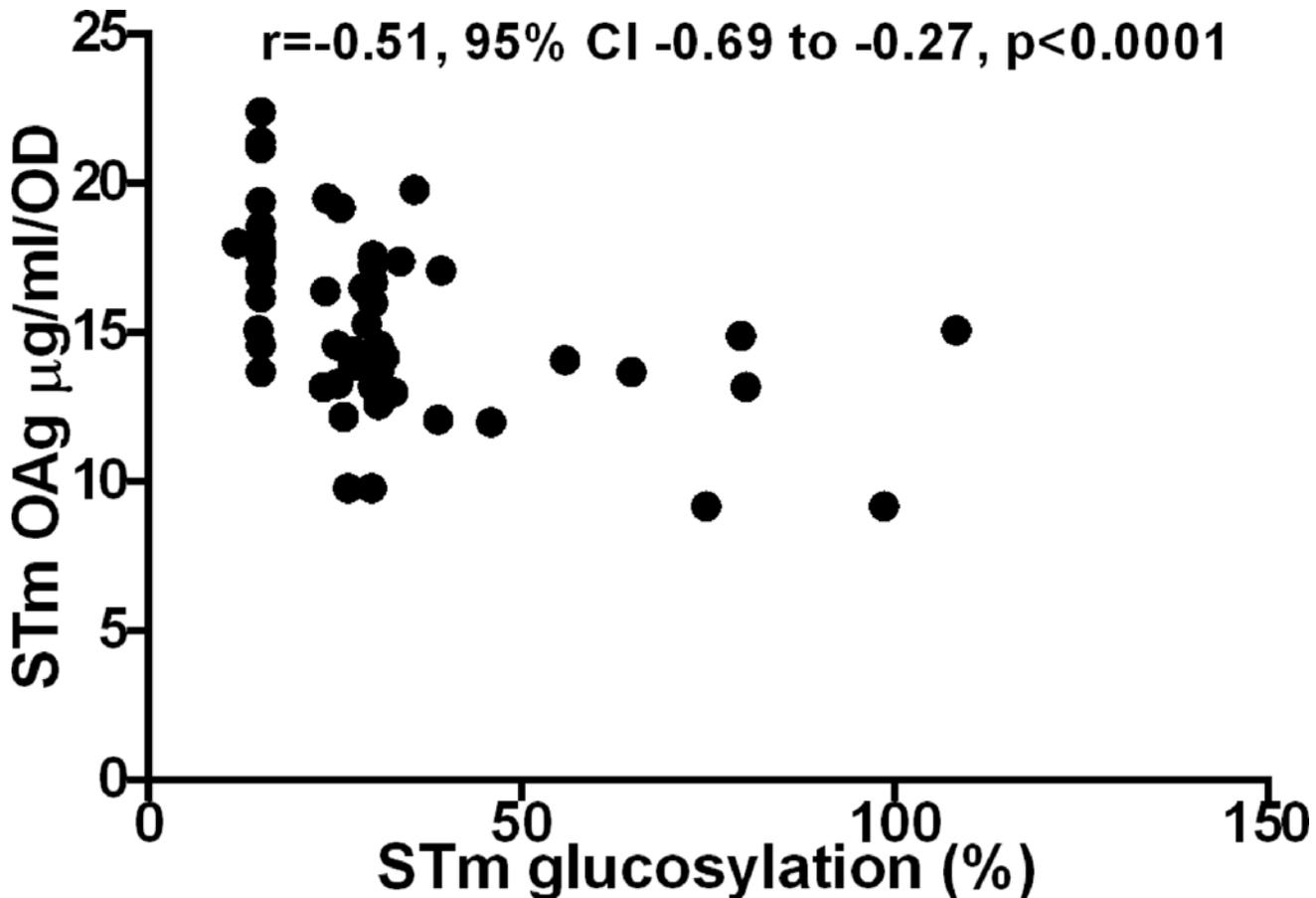


**Fig 5. Correlation of SBA results (a, c) and OAg production levels (b, d) in *S. Typhimurium* (a, b) and *S. Enteritidis* (c, d) isolates with clinical presentations.** Samples for which no clinical presentation was determined were excluded from analysis. Malawi serum pool was used for SBA. Invasive: *Salmonella* isolates from blood, urine, CSF. Gastrointestinal: *Salmonella* isolates from stools. Controls: *Salmonella* isolates from stools of healthy controls.

doi:10.1371/journal.pntd.0003573.g005

reported that O:12 glucosylation in Typhimurium is not constitutive, occurs during intracellular macrophage growth and is associated with enhanced intestinal colonization [46]. All *S. Typhimurium* isolates in our collection were glucosylated, with less than 30% of them at levels < 20%, showing that glucosylation is a very common feature among endemic strains. Similarly, 86% *S. Enteritidis* isolates investigated showed a glucosylation level >20%. Glucosylation in *S. Enteritidis* is not associated with specific OAg factors and it has not been extensively studied. A report on *S. Enteritidis* food-borne salmonellosis strains showed glucosylated LPS in some mouse isolates, closely related to strains obtained from eggs [47], but not in archived *S. Enteritidis* strains.

In *Shigella*, LPS glucosylation promotes bacterial invasion by enhancing type III secretion system (T3SS) function and is associated with reduced LPS chain length [48]. In *Salmonella*, T3SS is necessary for invasion of intestinal epithelium and survival within macrophages and further investigation into different glucosylation levels and T3SS-mediated invasion is still to be performed. The inverse correlation between *S. Typhimurium* OAg glucosylation and OAg expression observed in the current study may be related to T3SS function, as previously reported for *Shigella* [48] and *S. Typhimurium* [30]. *Salmonella* isolates defective for the synthesis of long and very long OAg species have been shown to have increased translocation of a



**Fig 6. Correlation of *S. Typhimurium* (STm) OAg production levels and glucosylation ( $p < 0.0001$ ).**  $r$  is the Spearman correlation coefficient. CI is confidence interval.

doi:10.1371/journal.pntd.0003573.g006

*Salmonella* pathogenicity island 1 (SPI1)-T3SS effector protein promoting invasion. Therefore, also for *Salmonella*, there may be the need to balance T3SS activity, possibly enhanced by shorter and glucosylated LPS, and protection against innate immune effectors, which is favoured by a thicker LPS layer [27,28].

The reasons for NTS disease presenting as invasive or enteric disease are not well understood and could relate to host immunity and/or pathogen virulence. Epidemiological data indicate a frequent presentation of the invasive syndrome, without diarrhoea symptoms, in younger age groups [49], often associated with other infections such as malaria and HIV [50–52]. However, specific investigations on the relationship between invasive and diarrheal disease are extremely limited, partially due to the limited microbiological resources in endemic African countries and few data are available on contemporaneous presence of NTS in stool and blood samples [53]. Another problem is that the concentration of *Salmonella* in the blood of bacteremic patients can be as low as 1 CFU/ml [54] and blood culture sensitivity is estimated to be about 50% [54].

The lack of obvious difference between phenotype of NTS isolates from invasive and gastrointestinal disease episodes is consistent with the concept that invasive NTS isolates are able to cause diarrhoea and vice versa. If true, this would implicate host immunity as the major determinant of NTS disease presentation in endemic sub-Saharan African countries. Other previous

studies [49] have not managed to identify specific pathogen determinants for either invasive or gastrointestinal syndromes, therefore the interplay between host and pathogen may be key.

There are several limitations to the current study: categorization of isolates as “invasive” or “gastroenteric” was based on clinical presentation, as confirmed by microbiological analysis. Blood and stool samples belonging to the same patient were rarely collected, making it impossible to exclude that bacteria from bacteremic patients were not also present in stool and vice versa [53]. Another limitation was the lack of patients’ outcome data, which could give further insights into possible differences in serovar- and isolate pathogenicity,

In summary, Kenyan NTS isolates from patients with bacteraemia and gastroenteritis were susceptible to antibody-mediated killing, supporting the development of an antibody-inducing vaccine against NTS for Africa. *S. Enteritidis* were generally less susceptible to killing than *S. Typhimurium* isolates and expressed higher levels of OAg, but while OAg expression correlated with antibody resistance for *S. Typhimurium* isolates, no correlation could be found for *S. Enteritidis*, supporting a role for other inherent bacterial factors in conferring resistance to antibody killing for this *Salmonella* serogroup. Serovar- and strain-specific differences in OAg expression and fine specificity were found. Those features could not be correlated to clinical presentation, so the same strains may be able to cause both invasive disease and diarrhoea. Whole genome analysis of invasive and gastrointestinal NTS isolates from Africa will provide additional insights regarding whether these strains are the same or different [53].

## Supporting Information

**S1 Table. a. List of *S. Typhimurium* study isolates and their available basic metadata; b. List of *S. Enteritidis* study isolates and their available basic metadata.**  
(XLSX)

**S2 Table. a. Serum bactericidal assay (SBA) results for *S. Typhimurium* isolates against pooled Malawian sera. b. Serum bactericidal assay (SBA) results for *S. Enteritidis* isolates against pooled Malawian sera.**  
(XLSX)

**S3 Table. a. O-acetylation results for *S. Typhimurium* and *S. Enteritidis* isolates. b. Glucosylation results for *S. Typhimurium* and *S. Enteritidis* isolates.**  
(XLSX)

## Acknowledgments

We thank the Directors, NVGH and KEMRI for permission to publish this work. We are grateful to Yunshan Goh for technical assistance in the laboratory.

## Author Contributions

Conceived and designed the experiments: RSO FM LL RA AS CAM SK SR. Performed the experiments: RSO FM LL RA SR. Analyzed the data: RSO FM LL RA CKO AWM GR AS SK CAM SR. Contributed reagents/materials/analysis tools: RSO FM LL RA CKO AWM GR AS SK CAM SR. Wrote the paper: RSO FM LL RA CKO AWM GR AS SK CAM SR.

## References

1. Feasey NA, Dougan G, Kingsley RA, Heyderman RS, Gordon MA (2012) Invasive non-typhoidal salmonella disease: an emerging and neglected tropical disease in Africa. *Lancet* 379: 2489–2499. doi: [10.1016/S0140-6736\(11\)61752-2](https://doi.org/10.1016/S0140-6736(11)61752-2) PMID: [22587967](https://pubmed.ncbi.nlm.nih.gov/22587967/)

2. MacLennan CA, Levine MM (2013) Invasive nontyphoidal *Salmonella* disease in Africa: current status. *Expert Rev Anti Infect Ther* 11: 443–446. doi: [10.1586/eri.13.27](https://doi.org/10.1586/eri.13.27) PMID: [23627848](https://pubmed.ncbi.nlm.nih.gov/23627848/)
3. Reddy EA, Shaw AV, Crump JA (2010) Community-acquired bloodstream infections in Africa: a systematic review and meta-analysis. *Lancet Infect Dis* 10: 417–432. doi: [10.1016/S1473-3099\(10\)70072-4](https://doi.org/10.1016/S1473-3099(10)70072-4) PMID: [20510282](https://pubmed.ncbi.nlm.nih.gov/20510282/)
4. Vieira A JA, Pires SM, Karlsmose S, Wegener HC, Wong DLF (2009) WHO Global Foodborne Infections Network Country Databank—a resource to link human and non-human sources of *Salmonella*. *Int Soc Vet Epidemiol Econ* 643: 512–517.
5. Langridge GC, Nair S, Wain J (2009) Nontyphoidal *Salmonella* serovars cause different degrees of invasive disease globally. *J Infect Dis* 199: 602–603. doi: [10.1086/596208](https://doi.org/10.1086/596208) PMID: [19281309](https://pubmed.ncbi.nlm.nih.gov/19281309/)
6. Mermin J, Lule J, Ekwaru JP, Downing R, Hughes P, et al. (2005) Cotrimoxazole prophylaxis by HIV-infected persons in Uganda reduces morbidity and mortality among HIV-uninfected family members. *AIDS* 19: 1035–1042. PMID: [15958834](https://pubmed.ncbi.nlm.nih.gov/15958834/)
7. Hohmann EL (2001) Nontyphoidal salmonellosis. *Clin Infect Dis* 32: 263–269. PMID: [11170916](https://pubmed.ncbi.nlm.nih.gov/11170916/)
8. Brent AJ, Oundo JO, Mwangi I, Ochola L, Lowe B, et al. (2006) *Salmonella* bacteremia in Kenyan children. *Pediatr Infect Dis J* 25: 230–236. PMID: [16511385](https://pubmed.ncbi.nlm.nih.gov/16511385/)
9. Gordon MA, Banda HT, Gondwe M, Gordon SB, Boeree MJ, et al. (2002) Non-typhoidal salmonella bacteraemia among HIV-infected Malawian adults: high mortality and frequent recrudescence. *AIDS* 16: 1633–1641. PMID: [12172085](https://pubmed.ncbi.nlm.nih.gov/12172085/)
10. Graham SM, Molyneux EM, Walsh AL, Cheesbrough JS, Molyneux ME, et al. (2000) Nontyphoidal *Salmonella* infections of children in tropical Africa. *Pediatr Infect Dis J* 19: 1189–1196. PMID: [11144383](https://pubmed.ncbi.nlm.nih.gov/11144383/)
11. Gordon MA, Graham SM, Walsh AL, Wilson L, Phiri A, et al. (2008) Epidemics of invasive *Salmonella enterica* serovar enteritidis and *S. enterica* Serovar typhimurium infection associated with multidrug resistance among adults and children in Malawi. *Clin Infect Dis* 46: 963–969. doi: [10.1086/529146](https://doi.org/10.1086/529146) PMID: [18444810](https://pubmed.ncbi.nlm.nih.gov/18444810/)
12. Kariuki S, Revathi G, Kariuki N, Kiiru J, Mwituria J, et al. (2006) Invasive multidrug-resistant nontyphoidal *Salmonella* infections in Africa: zoonotic or anthroponotic transmission? *J Med Microbiol* 55: 585–591. PMID: [16585646](https://pubmed.ncbi.nlm.nih.gov/16585646/)
13. Kingsley RA, Msefula CL, Thomson NR, Kariuki S, Holt KE, et al. (2009) Epidemic multiple drug resistant *Salmonella* Typhimurium causing invasive disease in sub-Saharan Africa have a distinct genotype. *Genome Res* 19: 2279–2287. doi: [10.1101/gr.091017.109](https://doi.org/10.1101/gr.091017.109) PMID: [19901036](https://pubmed.ncbi.nlm.nih.gov/19901036/)
14. Tabu C, Breiman RF, Ochieng B, Aura B, Cosmas L, et al. (2012) Differing burden and epidemiology of non-Typhi *Salmonella* bacteremia in rural and urban Kenya, 2006–2009. *PLoS One* 7: e31237. doi: [10.1371/journal.pone.0031237](https://doi.org/10.1371/journal.pone.0031237) PMID: [22363591](https://pubmed.ncbi.nlm.nih.gov/22363591/)
15. Talbert AW, Mwaniki M, Mwarumba S, Newton CR, Berkley JA (2010) Invasive bacterial infections in neonates and young infants born outside hospital admitted to a rural hospital in Kenya. *Pediatr Infect Dis J* 29: 945–949. doi: [10.1097/INF.0b013e3181dfca8c](https://doi.org/10.1097/INF.0b013e3181dfca8c) PMID: [20418799](https://pubmed.ncbi.nlm.nih.gov/20418799/)
16. O'Reilly CE, Jaron P, Ochieng B, Nyaguara A, Tate JE, et al. (2012) Risk factors for death among children less than 5 years old hospitalized with diarrhea in rural western Kenya, 2005–2007: a cohort study. *PLoS Med* 9: e1001256. doi: [10.1371/journal.pmed.1001256](https://doi.org/10.1371/journal.pmed.1001256) PMID: [22802736](https://pubmed.ncbi.nlm.nih.gov/22802736/)
17. Okoro CK, Kingsley RA, Connor TR, Harris SR, Parry CM, et al. (2012) Intracontinental spread of human invasive *Salmonella* Typhimurium pathovariants in sub-Saharan Africa. *Nat Genet* 44: 1215–1221. doi: [10.1038/ng.2423](https://doi.org/10.1038/ng.2423) PMID: [23023330](https://pubmed.ncbi.nlm.nih.gov/23023330/)
18. Ikumapayi UN, Antonio M, Sonne-Hansen J, Biney E, Enwere G, et al. (2007) Molecular epidemiology of community-acquired invasive non-typhoidal *Salmonella* among children aged 2–29 months in rural Gambia and discovery of a new serovar, *Salmonella enterica* Dingiri. *J Med Microbiol* 56: 1479–1484. PMID: [17965348](https://pubmed.ncbi.nlm.nih.gov/17965348/)
19. Paglietti B, Falchi G, Mason P, Chitsatso O, Nair S, et al. (2013) Diversity among human non-typhoidal salmonellae isolates from Zimbabwe. *Trans R Soc Trop Med Hyg* 107: 487–492. doi: [10.1093/trstmh/trt046](https://doi.org/10.1093/trstmh/trt046) PMID: [23843558](https://pubmed.ncbi.nlm.nih.gov/23843558/)
20. Galanis E, Lo Fo Wong DM, Patrick ME, Binsztein N, Cieslik A, et al. (2006) Web-based surveillance and global *Salmonella* distribution, 2000–2002. *Emerg Infect Dis* 12: 381–388. PMID: [16704773](https://pubmed.ncbi.nlm.nih.gov/16704773/)
21. Rietschel ET, Brade L., Linder B., and Zaehring U., editor (1992) *Bacterial endotoxic lipopolysaccharides*. Boca Raton, Florida: CRC Press, Incorporation. 3–41 p.
22. Park PH, Huang H, McMullen MR, Mandal P, Sun L, et al. (2008) Suppression of lipopolysaccharide-stimulated tumor necrosis factor- $\alpha$  production by adiponectin is mediated by transcriptional and post-transcriptional mechanisms. *J Biol Chem* 283: 26850–26858. doi: [10.1074/jbc.M802787200](https://doi.org/10.1074/jbc.M802787200) PMID: [18678874](https://pubmed.ncbi.nlm.nih.gov/18678874/)

23. Raetz CR, Whitfield C (2002) Lipopolysaccharide endotoxins. *Annu Rev Biochem* 71: 635–700. PMID: [12045108](#)
24. Joiner KA, Hammer CH, Brown EJ, Cole RJ, Frank MM (1982) Studies on the mechanism of bacterial resistance to complement-mediated killing. I. Terminal complement components are deposited and released from *Salmonella minnesota* S218 without causing bacterial death. *J Exp Med* 155: 797–808. PMID: [6801179](#)
25. Saxen H, Reima I, Makela PH (1987) Alternative complement pathway activation by *Salmonella* O polysaccharide as a virulence determinant in the mouse. *Microb Pathog* 2: 15–28. PMID: [3333794](#)
26. MacLennan CA, Gondwe EN, Msefula CL, Kingsley RA, Thomson NR, et al. (2008) The neglected role of antibody in protection against bacteremia caused by nontyphoidal strains of *Salmonella* in African children. *J Clin Invest* 118: 1553–1562. doi: [10.1172/JCI33998](#) PMID: [18357343](#)
27. Joiner KA, Grossman N, Schmetz M, Leive L (1986) C3 binds preferentially to long-chain lipopolysaccharide during alternative pathway activation by *Salmonella montevideo*. *J Immunol* 136: 710–715. PMID: [2416822](#)
28. Murray GL, Attridge SR, Morona R (2006) Altering the length of the lipopolysaccharide O antigen has an impact on the interaction of *Salmonella enterica* serovar Typhimurium with macrophages and complement. *J Bacteriol* 188: 2735–2739. PMID: [16547065](#)
29. Bravo D, Silva C, Carter JA, Hoare A, Alvarez SA, et al. (2008) Growth-phase regulation of lipopolysaccharide O-antigen chain length influences serum resistance in serovars of *Salmonella*. *J Med Microbiol* 57: 938–946. doi: [10.1099/jmm.0.47848-0](#) PMID: [18628492](#)
30. Holzer SU, Schlumberger MC, Jackel D, Hensel M (2009) Effect of the O-antigen length of lipopolysaccharide on the functions of Type III secretion systems in *Salmonella enterica*. *Infect Immun* 77: 5458–5470. doi: [10.1128/IAI.00871-09](#) PMID: [19797066](#)
31. Goh YS, MacLennan CA (2013) Invasive African nontyphoidal *Salmonella* requires high levels of complement for cell-free antibody-dependent killing. *J Immunol Methods* 387: 121–129. doi: [10.1016/j.jim.2012.10.005](#) PMID: [23085530](#)
32. MacLennan CA, Gilchrist JJ, Gordon MA, Cunningham AF, Cobbold M, et al. (2010) Dysregulated humoral immunity to nontyphoidal *Salmonella* in HIV-infected African adults. *Science* 328: 508–512. doi: [10.1126/science.1180346](#) PMID: [20413503](#)
33. Rondini S, Lanzilao L, Necchi F, O'Shaughnessy CM, Micoli F, et al. (2013) Invasive African *Salmonella* Typhimurium induces bactericidal antibodies against O-antigens. *Microb Pathog* 63: 19–23. doi: [10.1016/j.micpath.2013.05.014](#) PMID: [23756206](#)
34. Micoli F, Rondini S, Gavini M, Pisoni I, Lanzilao L, et al. (2013) A scalable method for O-antigen purification applied to various *Salmonella* serovars. *Anal Biochem* 434: 136–145. doi: [10.1016/j.ab.2012.10.038](#) PMID: [23142430](#)
35. Dubois M, Gilles R.A., Hamilton J.K., Roberts P.A. & Smith F. (1956) Colorimetric method for determination of sugar and related substances. *Anal Chem* 28: 350–356.
36. Micoli F, Rondini S, Gavini M, Lanzilao L, Medagliani D, et al. (2012) O:2-CRM(197) conjugates against *Salmonella Paratyphi* A. *PLoS One* 7: e47039. doi: [10.1371/journal.pone.0047039](#) PMID: [23144798](#)
37. Kotloff KL, Nataro JP, Blackwelder WC, Nasrin D, Farag TH, et al. (2013) Burden and aetiology of diarrhoeal disease in infants and young children in developing countries (the Global Enteric Multicenter Study, GEMS): a prospective, case-control study. *Lancet* 382: 209–222. doi: [10.1016/S0140-6736\(13\)60844-2](#) PMID: [23680352](#)
38. Nyirenda TS, Gilchrist JJ, Feasey NA, Glennie SJ, Bar-Zeev N, et al. (2014) Sequential Acquisition of T Cells and Antibodies to Nontyphoidal *Salmonella* in Malawian Children. *J Infect Dis*.
39. O'Shaughnessy CM, Cunningham AF, MacLennan CA (2012) The stability of complement-mediated bactericidal activity in human serum against *Salmonella*. *PLoS One* 7: e49147. doi: [10.1371/journal.pone.0049147](#) PMID: [23145102](#)
40. Dhanoa A, Fatt QK (2009) Non-typhoidal *Salmonella* bacteraemia: epidemiology, clinical characteristics and its' association with severe immunosuppression. *Ann Clin Microbiol Antimicrob* 8: 15. doi: [10.1186/1476-0711-8-15](#) PMID: [19445730](#)
41. Moller AS, Ovstebø R, Westvik AB, Joo GB, Haug KB, et al. (2003) Effects of bacterial cell wall components (PAMPs) on the expression of monocyte chemoattractant protein-1 (MCP-1), macrophage inflammatory protein-1alpha (MIP-1alpha) and the chemokine receptor CCR2 by purified human blood monocytes. *J Endotoxin Res* 9: 349–360. PMID: [14733721](#)
42. Nevola JJ, Laux DC, Cohen PS (1987) In vivo colonization of the mouse large intestine and in vitro penetration of intestinal mucus by an avirulent smooth strain of *Salmonella typhimurium* and its lipopolysaccharide-deficient mutant. *Infect Immun* 55: 2884–2890. PMID: [3316026](#)

43. Wollin R, Stocker BA, Lindberg AA (1987) Lysogenic conversion of *Salmonella typhimurium* bacteriophages A3 and A4 consists of O-acetylation of rhamnose of the repeating unit of the O-antigenic polysaccharide chain. *J Bacteriol* 169: 1003–1009. PMID: [3546260](#)
44. Micoli F, Ravenscroft N, Cescutti P, Stefanetti G, Londero S, et al. (2014) Structural analysis of O-polysaccharide chains extracted from different *Salmonella Typhimurium* strains. *Carbohydr Res* 385: 1–8. doi: [10.1016/j.carres.2013.12.003](#) PMID: [24384528](#)
45. Makela PH (1973) Glucosylation of lipopolysaccharide in *Salmonella*: mutants negative for O antigen factor 1221. *J Bacteriol* 116: 847–856. PMID: [4583253](#)
46. Bogomolnaya LM, Santiviago CA, Yang HJ, Baumler AJ, Andrews-Polymenis HL (2008) 'Form variation' of the O12 antigen is critical for persistence of *Salmonella Typhimurium* in the murine intestine. *Mol Microbiol* 70: 1105–1119. doi: [10.1111/j.1365-2958.2008.06461.x](#) PMID: [18826410](#)
47. Parker CT, Liebana E, Henzler DJ, Guard-Petter J (2001) Lipopolysaccharide O-chain microheterogeneity of *Salmonella* serotypes Enteritidis and Typhimurium. *Environ Microbiol* 3: 332–342. PMID: [11422320](#)
48. West NP, Sansonetti P, Mounier J, Exley RM, Parsot C, et al. (2005) Optimization of virulence functions through glucosylation of *Shigella* LPS. *Science* 307: 1313–1317. PMID: [15731456](#)
49. Kariuki S, Revathi G, Kariuki N, Kiiru J, Mwituria J, et al. (2006) Characterisation of community acquired non-typhoidal *Salmonella* from bacteraemia and diarrhoeal infections in children admitted to hospital in Nairobi, Kenya. *BMC Microbiol* 6: 101. PMID: [17173674](#)
50. MacLennan CA (2012) Host defense against malaria favors *Salmonella*. *Nat Med* 18: 21–22. doi: [10.1038/nm.2636](#) PMID: [22227659](#)
51. Gordon MA (2008) *Salmonella* infections in immunocompromised adults. *J Infect* 56: 413–422. doi: [10.1016/j.jinf.2008.03.012](#) PMID: [18474400](#)
52. Biggs HM, Lester R, Nadjm B, Mtove G, Todd JE, et al. (2014) Invasive *Salmonella* infections in areas of high and low malaria transmission intensity in Tanzania. *Clin Infect Dis* 58: 638–647. doi: [10.1093/cid/cit798](#) PMID: [24336909](#)
53. Onsare RS, MacLennan CA (2014) New genomics studies to understand the link between nontyphoidal *Salmonella* bacteremia and gastroenteritis in Africa. *J Infect Dev Ctries* 8: 252–253. doi: [10.3855/jidc.4377](#) PMID: [24518640](#)
54. Tennant SM, Zhang Y, Galen JE, Geddes CD, Levine MM (2011) Ultra-fast and sensitive detection of non-typhoidal *Salmonella* using microwave-accelerated metal-enhanced fluorescence ("MAMEF"). *PLoS One* 6: e18700. doi: [10.1371/journal.pone.0018700](#) PMID: [21494634](#)